

Ultrastructural evaluation of compression wood-like properties of common juniper (*Juniperus communis* L.)

Tuomas Hänninen^{1,*}, Pekka Tukiainen¹,
Kirsi Svedström², Ritva Serimaa², Pekka Saranpää³,
Eero Kontturi¹, Mark Hughes¹ and Tapani Vuorinen¹

¹Department of Forest Products Technology, School of Chemical Technology, Aalto University, Aalto, Finland

²Department of Physics, Division of Materials Physics, University of Helsinki, Helsinki, Finland

³Finnish Forest Research Institute Metla, Vantaa, Finland

*Corresponding author.

Department of Forest Products Technology,
School of Chemical Technology,
Aalto University, P.O. Box 16300,
FI-00076 Aalto, Finland
Phone: +358-50-512-4223
E-mail: tuomas.hanninen@aalto.fi

Abstract

To date, very little is known about the ultrastructure of common juniper, *Juniperus communis* L. (juniper). In this study, the mechanical properties of juniper, its chemistry and ultrastructure has been analyzed. The data are presented in comparison to the normal wood (NW) and compression wood (CW) of spruce. Helical grooves, which are a characteristic of CW, were clearly visible in micrographs of juniper. The angle of the grooves with respect to the fiber axis was ca. 40°, which correlates with the microfibril angle determined by X-ray diffraction. Data from 4-point bending tests show that juniper and spruce CW exhibited similar behavior. The elastic moduli of both species were only ca. half from that of NW spruce. The composition of juniper fibers resembled that of CW fiber with respect to high lignin and hemicelluloses contents. However, the galactose content in CW of juniper was low and in CW of spruce was high. Raman imaging clearly revealed that the lignin/cellulose ratio in the fiber cell wall of juniper was similar to that of NW spruce.

Keywords: *Juniperus communis* L.; mechanical properties; microfibril angle (MFA); Raman imaging.

Introduction

Trees respond to stimuli – such as inclination of the ground, strong wind, or heavy snow on the branches – by forming reaction wood. Hardwoods develop the so-called tension wood in an attempt to pull the stem (or branches) upright, whereas softwood forms compression wood (CW) to generate compressive stress. CW differs from normal wood (NW) in its chemistry,

mechanical properties, and the structure of the fibers (Timell 1986). The formation, physical and chemical characteristics of reaction wood are still a matter of scientific interest as demonstrated in the literature of the past decade. For example, the following topics have been the focus of attention: mechanical properties (Gindl 2002; Gindl and Teischinger 2003), air permeability (Tarmian and Perré 2009), wood-water relationship (Perré 2007), viscoelastic properties (Placet et al. 2007), morphological and chemical properties (Önnerud 2003; Yeh et al. 2005, 2006; Altaner et al. 2007; Tokareva et al. 2007; Kibblewhite et al. 2010), and quantitative chemical indicators to assess the gradation of CW (Nanayakkara et al. 2009). One of the distinct features of CW is the presence of the S_{2L} layer, which is within the S2 cell wall containing a high concentration of lignin (Timell 1986; Donaldson et al. 1999; Gierlinger et al. 2010). CW cells are also known to lack the S3 layer and have helical grooves in the S2 layer (Timell 1982).

The wood of the genus *Juniperus* has many CW-like features. Common juniper (*Juniperus communis* L.), referred to in this work as juniper, is widely distributed in the northern hemisphere. It is known for its berries, which gives the flavor to gin, strong scented extractives, and resilient mechanical properties (Eckenwalder 2009). The papermaking properties of pulped juniper were reported by Hänninen et al. (2011a) and the fiber dimensions of juniper resemble those of CW from other softwood species. Indeed, the juniper fibers were shorter and their cell wall was thicker, which is commonplace with CW fibers. Compared with the NW of other softwoods, a high microfibril angle (MFA), which is a common feature of CW, is an attribute for juniper (Kantola and Seitsonen 1961; Kantola and Kähkönen 1963). In addition, rounded tracheids and intercellular spaces are typical for the NW of the genus *Juniperus*, which are commonly characteristics of CW (Timell 1983). The mechanical properties of juniper bear a closer resemblance to those of CW than to NW spruce. It is intriguing that the NW in one species should possess characteristics that are so close to CW.

The aim of the present study was to investigate morphological and chemical peculiarities of juniper in comparison to spruce CW and NW. The ultrastructural details were analyzed by light microscopy, Raman imaging, and X-ray diffraction. In addition, the chemical composition and mechanical properties were determined.

Materials and methods

Norway spruce (*Picea abies* [L.] Karst.) was obtained from Mikkeli, south-eastern Finland. Samples were cut from the same stem; NW being taken from approximately breast height and CW lower in the stem, close to the stump. Stems of common juniper (*Juniperus*

communis L.) were collected from Solböle, south-western Finland. Samples were cut from the same, approximately 7 cm thick, straight and branchless stem roughly at a height of 50 cm. This material does not contain juvenile wood (JW).

Radial sections 16 μm in thickness were cut with a cryomicrotome at -14°C . Sections were stained with safranin-Alcian blue and the helical grooves or cavities were observed under a Olympus BX60 microscope connected to Spot insight B/W video camera (Diagnostic Instruments, Inc., USA) and Image-Pro plus for Windows program. Images of radial sections were captured at 20 \times magnification.

Static bending tests

Juniper, spruce NW and CW were cut to small rectangular beams with dimensions 3 mm (r), 6 mm (t), and 60 mm (l). The specimens were conditioned to constant weight in a climate chamber (RH 65% and T 20°C) for ca. 2 weeks prior to testing. The annual ring widths in juniper were smaller than those in spruce NW and CW, which resulted in a different number of annual rings in the specimens. This gave rise to differences in the amounts of late wood (LW) different samples, which may have affected the stiffness of the specimens.

The mechanical properties were determined by a four-point bending test with an outer span of 37.5 mm and an inner span of 12.5 mm (Figure 1). Testing was conducted on a small loading device (stepper motor driven) designed for the use with a light microscope (Wild MZ8, Leica, Wetzlar, Germany). Force was measured with a 500 N load cell (U9B, HBM, Darmstadt, Germany). Loading was conducted under displacement control at a cross-head displacement rate of 0.5 mm min^{-1} . Specimens were loaded to 50 N and then unloaded to 5 N prior to the actual measurement, where samples were loaded to failure. During testing, load and cross-head displacement data were recorded (Solartron AX/2.5/SH, Solartron metrology, West Sussex, UK) and the specimen observed with an optical microscope equipped with a CCD camera (C-Mount 3-CCD, JVC, Japan). Micrographs were captured at 5 N intervals.

The longitudinal modulus of elasticity (MOE) was calculated from classical beam theory for homogeneous beams of rectangular cross-section. Total cross-sectional area was used. MOE was calculated from the crosshead displacement (assumed to be equal to the deflection of the beam under the loading head) and load data between 0.1 and 0.4 of the maximum load. The effect of shear forces and indentation of the supports and loading head were neglected.

$$MOE_B = \frac{5}{27} \cdot \frac{L^3}{T \cdot R^3} \cdot \frac{\Delta F}{\Delta f} \quad (1)$$

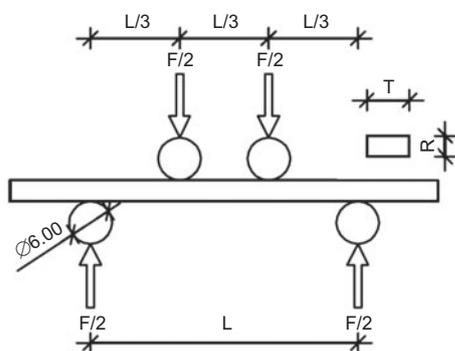


Figure 1 The 4-point bend test set-up; L is outer span, $L/3$ is inner span, R is specimen height, and T is specimen width.

where MOE_B is modulus of elasticity determined in 4-point bending, L is the outer span, T is the specimen width, R is the specimen height, ΔF is the load difference, and Δf is the deflection under the loading head (Gere 1997).

Chemical composition

Wood samples were milled in a Wiley mill and extracted with acetone (AnalaR NORMAPUR, VWR, Fontenay-sous-Bois, France) to determine the extractives content. The Klason lignin and carbohydrate contents were determined based on acid hydrolysis according to a method detailed by Sluiter et al. (2008). The monosaccharides in the resulting hydrolysates were quantified by high-performance anion-exchange chromatography (HPAEC) (Dionex ICS-3000, CarboPac PA20 column, pulsed amperometric detection, PAD).

X-ray diffraction (XRD)

For XRD, all samples were cut tangentially from the stems with a scalpel to a (radial) thickness of ca. 1 mm, dimensions: 10 (l) \times 10 (t) mm^2 . Two samples were cut from the juniper stem in the middle between the pith and bark (1 mm thick); the pieces included several annual rings. The spruce NW samples were cut from the earlywood (EW) of the 16th and the 19th annual rings, and the CW samples were cut from the EW of the 20th, the 21st, and the 22nd annual rings.

The MFA distributions were determined from the azimuthal intensity profiles of the cellulose reflections 200 and 004 measured in symmetrical transmission mode by means of the set-up with the 4-circle goniometer as described by Hänninen et al. (2011a). The MFAs were determined from the intensity profiles by fitting pairs of Gaussian functions into the profiles as detailed by Sarén et al. (2001).

Diffraction patterns were measured with the same X-ray set-up for determining the dimensions of cellulose crystallites. The crystal length was determined from the reflection 004 measured in symmetrical transmission mode, and the crystal width was determined from the reflection 200 measured in symmetrical reflection mode. The crystal dimensions were obtained from the widths and positions of the reflections by the Scherrer equation (Andersson et al. 2003). The azimuthal intensity profiles of the four juniper samples (Hänninen et al. 2011a) were also analyzed here at high angles (close to 90°).

Raman imaging

Sample preparation and Raman imaging were performed according to Hänninen et al. (2011b). The samples were analyzed with an $\alpha 300$ R Confocal Raman microscope (Witec GmbH, Germany, www.witec.de) at ambient conditions. The instrument works with frequency doubled Nd: YAG laser (532.35 nm, 10 mW) and a Nikon 100 \times (NA=0.95) air objective, and it was equipped with a DU970N-BV EMCCD camera behind a 600-line mm^{-1} grating. The excitation laser was polarized horizontally. The integration times and measured areas differed from image to image depending on how prone the samples were to burning during measurement. Only the LW cells could be analyzed as the resolution of Raman imaging is ca. 300 nm due to the diffraction limit and thus the features of thin walled EW cells would certainly be indistinguishable.

Results and discussion

Micrographs of Norway spruce CW and juniper are presented in Figure 2. Helical grooves or cavities characteristic to CW (Timell 1986) were clearly visible on juniper micrographs

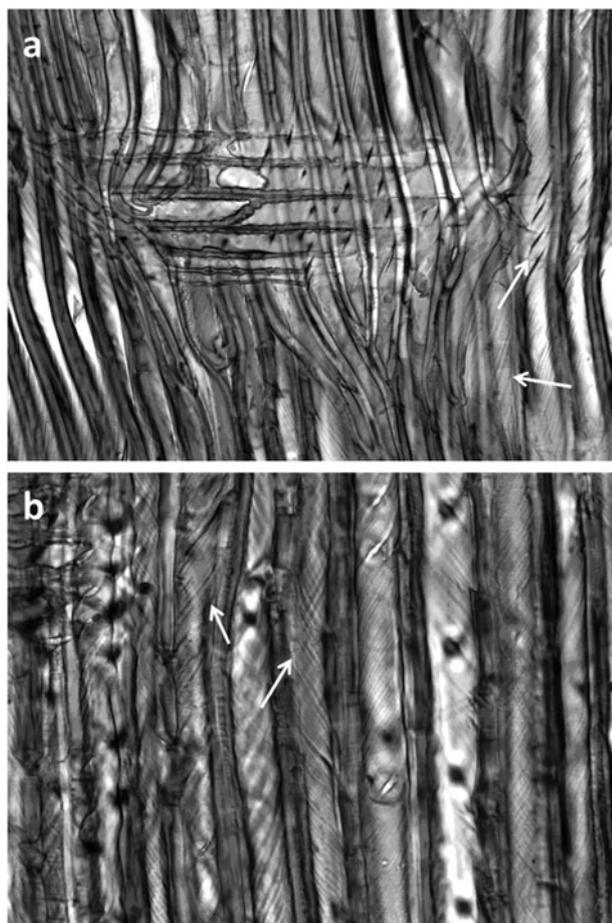


Figure 2 Radial sections of (a) compression wood of spruce and (b) common juniper. Helical grooves or cavities indicating high MFA of the cell wall are clearly visible. Some of the defects are indicated with arrows.

(Figure 2b). The angle of the grooves with respect to the fiber axis is ca. 40–45°. This correlates well with the MFA data from XRD (Table 1).

The mean MFA measured by XRD of both spruce CW and juniper was around 35° (Table 1), which corresponds to literature values. However, a very large variation in MFA (24.7–37°) was reported for juniper (Kantola and Seitsonen 1961; Kantola and Kähkönen 1963; Timell 1986; Burgert et al. 2004). Reportedly, MFA values for the juniper CW are very close to the MFA values in the juniper NW (Kantola and Seitsonen 1961), and this is the reason why the presence of CW cannot be entirely excluded.

The mean and standard deviation of the MFA given in Table 1 correspond to the largest peak observed in the azimuthal intensity profiles of the samples (Figure 3). It was assumed that this peak arises from the S2 layer of the cell wall, because it is the thickest. From the azimuthal intensity profiles of juniper and spruce CW contribution, other than from the S2 cell wall, could be distinguished (peak 1 in Figure 3). The contribution was assumed to be from the S1 layer, as CW lacks the S3 layer and the arrangement of the fibrils in the S1 is thought to be random. The mean MFAs in the S1 layer were 84–89°

Table 1 The mean microfibril angle (MFA) of the S2 cell wall layer, the crystal length and width determined by XRD.

Sample	MFA (°)	Crystal length (nm)	Crystal width (nm)
Spruce	0.2 (7)	40±5	3.08±0.05
Spruce CW	35 (8)	20±2	3.06±0.05
Juniper	35 (9)	21±2	2.92±0.05

The standard deviation of the MFA values is presented within parentheses. The error margins for the dimensions of the crystallites are based on the accuracy of the measurement and analysis.

and 66–90° for spruce CW and juniper, respectively, which are in agreement with literature data, where an almost perpendicular orientation to the fiber axis was found (Brändström 2001). The ratio of the integrated intensities of peak 1 (I_1) to that of peaks 1 and 2 (I_1+I_2) was 17%–20% for spruce CW and 11%–41% for juniper. If the peaks 1 and 2 correspond to the S1 and the S2 as assumed, then this ratio corresponds to the thickness ratio of the layers.

The crystal length is the same in juniper and spruce CW (ca. 20 nm), whereas in spruce NW it was twice as high (ca. 40 nm). A shorter crystal length in CW was found by Tanaka et al. (2005). Shorter crystal lengths result in higher elasticity of the MF because the number of dissipative amorphous areas is increased (Page 1983). Crystals in juniper wood (Table 1) were longer and thinner than in juniper pulp (Hänninen et al. 2011a). Similar changes in the crystal dimensions during pulping of spruce and birch wood were discussed in detail by Leppänen et al. (2010). NW and CW of spruce had the same crystal width, whereas it was slightly smaller in juniper. The crystal widths determined from the cellulose reflection 200 are presented in Figure 4. All crystal widths are in agreement with literature values (Andersson et al. 2003; Davidson et al. 2004; Yanagisawa et al. 2005; McLean et al. 2010).

MFA and density belong to the most important factors affecting the mechanical properties of wood. An increase in density elevates the stiffness of the material due to the greater

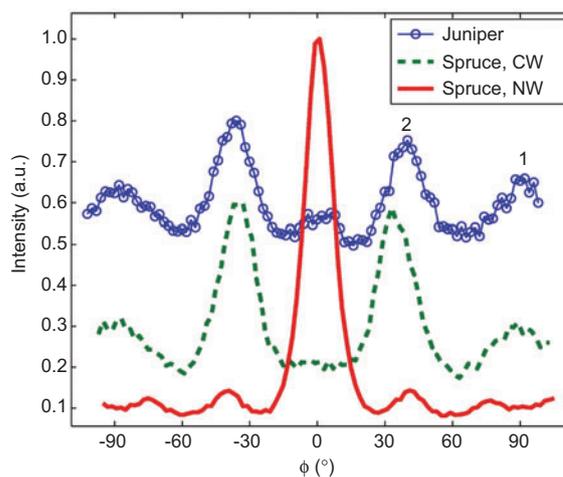


Figure 3 Azimuthal intensity profiles of the cellulose reflection 004 for juniper, spruce compression wood (CW), and spruce normal wood (NW).

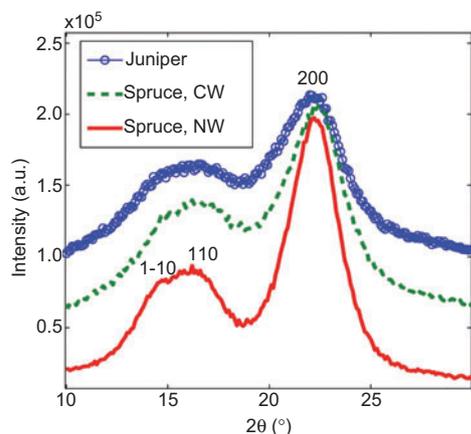


Figure 4 Radial intensity profile of the cellulose reflection 200 for juniper, spruce compression wood (CW), and spruce normal wood (NW), from which the cellulose crystal widths were determined.

volume of cell wall material. By contrast, increasing MFA leads to decreasing stiffness (Evans and Ilic 2001; Salmén and Fahlén 2009; McLean et al. 2010). The elastic moduli of the samples with high MFA (Table 2) – in juniper and spruce CW – were clearly lower than that of NW spruce. Although CW and juniper were significantly denser than spruce NW, the effect of MFA is clearly reflected in the data of elastic modulus. The specific stiffness data of NW spruce (MOE/ρ or MOE/ρ^2 , where ρ is density) are more than twice as high than those of spruce CW or juniper. Similar differences between spruce and yew, the MFA and density data of which are also very high, were already observed by Keunecke et al. (2008).

Table 2 Elastic moduli (MOE), densities, and maximum stresses at break values for spruce, spruce compression wood (CW), and juniper samples.

Sample	Density (kg m ⁻³)	MOE (MPa)	Bending strength (MPa)
Spruce	415	8215	72
	455	7944	68
	529	7693	76
	440	9583	82
	428	9591	75
	454±45	8605±915	74±5
Spruce CW	600	5116	92
	572	4789	95
	564	4514	90
	563	4743	93
	574	5646	97
	575±15	4962±439	93±3
Juniper	809	4709	113
	578	5344	84
	736	3869	104
	575	4906	80
	539	5153	81
	647±118	4796±572	92±15

Values from five parallel samples are shown together with the averages (in bold font).

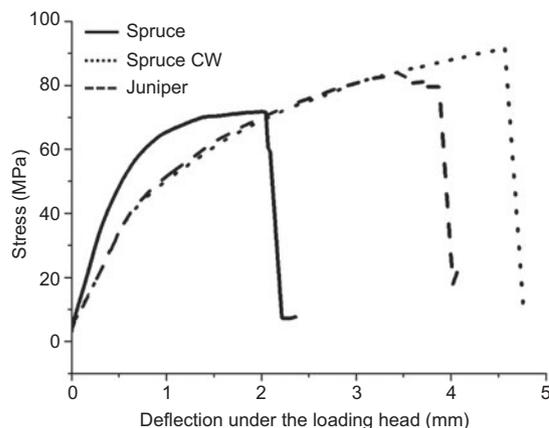


Figure 5 Stress versus deflection under the loading head curves of juniper, spruce, and spruce compression wood.

Spruce CW and juniper both display slightly higher bending strengths than the spruce NW. When density is taken into account, the specific strength of all groups is comparable. Generally, MFA has a strong effect on the tensile strength, with higher MFAs leading to lower tensile strength (Page et al. 1972). By contrast, compression strength rises almost linearly with density in air-dried wood. In bending, both tensile and compressive stresses are present and thus it may be presumed that the competing effects cancel each other. Nevertheless, differences in the fracture behavior were observed. As expected, in all juniper and spruce CW specimens failure began on the side under tension, whereas in the NW of spruce both tension and compression failures occurred.

Typical stress-deflection histories are presented in Figure 5. As is visible, the deflection of juniper and spruce CW at failure is much larger than that of the spruce NW. By comparing the areas under the stress-deflection curves it is obvious that juniper and spruce CW are able to dissipate significantly greater amounts of energy prior to fracture.

The Klason lignin content of spruce NW is significantly lower than in CW, as expected (Timell 1986; Yeh et al. 2006). The lignin content of juniper is higher than that of spruce but it is lower than in CW and corresponds well with the lignin content of juniper reported earlier (Timell 1986). The samples with higher lignin content also possess higher density. Juniper and spruce CW containing high lignin contents are significantly denser than spruce NW. The extractives content of juniper is significantly greater than that of spruce NW and spruce CW, whereas the latter has slightly elevated extractives content compared to spruce NW (Table 3).

Table 3 Lignin and extractives contents (% of wood) of spruce, spruce compression wood and juniper.

Sample	Lignin (%)		Extractives (%)
	Acid soluble	Klason	
Spruce	0.5	28.9	0.9
Spruce CW	0.5	36.6	1.7
Juniper	0.7	30.1	5.8

Table 4 Carbohydrate composition (%) in acid hydrolysates of spruce, spruce compression wood (CW), and juniper.

Sugar	Spruce		Juniper (%)
	NW (%)	CW (%)	
Ara	1.3	1.1	1.4
Rha	0.3	0.4	0.5
Gal	5.5	17.5	7.4
Glc	67.7	59.5	61.9
Xyl	9.0	10.4	11.9
Man	16.3	11.2	17.1

Ara, arabinose; Rha, rhamnose; Gal, galactose; Glc, glucose; Xyl, xylose; Man, mannose.

The spruce CW and juniper contain higher amounts of non-cellulosic carbohydrates than the spruce NW (Table 4). This is a known characteristic difference between NW and CW (Timell 1986). Spruce CW contains a large amount of galactose originating from (1–4)- β -galactan which is

reportedly higher in CW (Yeh et al. 2006; Mast et al. 2009). This type of polysaccharide is situated mainly in the S2_(L) layer in particular (Altaner et al. 2007, 2010). Slightly elevated xylose contents were also observed in juniper and spruce CW.

Raman images (Figure 6) show an even lignin/cellulose distribution in juniper, whereas the lignin content increases drastically in the primary cell wall (S1) and in the composed middle lamella (CML). A similar distribution was observed for *Juniper excelsa* (Adamopoulos and Koch 2011) by UMSP, although the resolution of this instrument is not high enough to distinguish clearly between small details in the cell wall. Raman images illustrating the lignin and lignin/cellulose distributions in the cell walls (Figure 6) resemble previously published ones (Gierlinger et al. 2010). The lignin/cellulose distribution of juniper is similar to that of spruce NW, whereas the additional lignin layer S2_(L) can only be observed in the CW samples. No differences in distribution could be detected between the images taken from different growth rings in any of the samples.

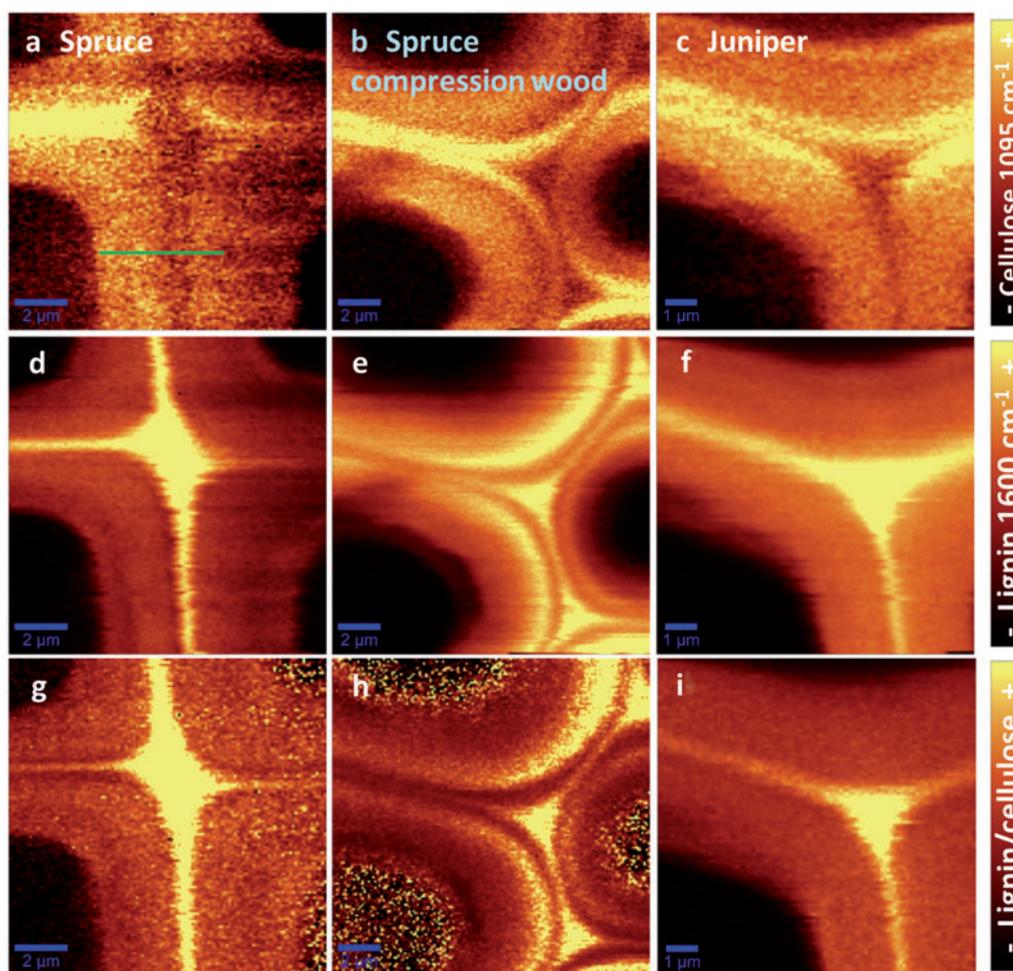


Figure 6 Raman images of spruce, spruce compression wood, and juniper. Images are constructed according to area of characteristic Raman bands of cellulose (a–c), lignin (d–f), and lignin/cellulose ratio (g–i). Laser polarization direction is indicated with green line. Scale in lignin/cellulose ratio images of spruce and juniper (g and i) images ranges from 0 to 6 arbitrary units and for spruce compression wood (h) from 0 to 12 arbitrary units.

Measurements by polarized laser in Raman spectroscopy enable the detection of S1 cell wall because the Raman spectrum of cellulose is sensitive to orientation (Atalla et al. 1980; Wiley and Atalla 1987; Gierlinger et al. 2010). The S1 layer is aligned along the plane of polarization of the laser (horizontal cell walls layer) and appears bright in the Raman images based on the characteristic cellulose band at 1095 cm^{-1} (Figure 6a–c). The bright color indicating larger area for this Raman band is a feature arising from the orientation of cellulose in the S1 layer. Therefore, the intensity of Raman scattering does not correlate with the concentration of cellulose in this case. This feature, however, is useful for distinguishing the S1 layer from the S2. No noticeable difference could be observed in the thickness of the S1 layers, which differs from the MFA determination by XRD. In XRD data, contribution of another cell wall layer with very low MFA could be distinguished, which is probably due to thick S1 layers. The poor resolution of Raman imaging and the fact that only LW cells were analyzed may explain the finding that S1 thicknesses could not be differentiated.

Conclusions

Literature data about *J. communis* could be confirmed and specified concerning its CW-like features, such as helical grooves, fiber dimensions (Hänninen et al. 2011a), high elasticity, and high MFA. The helical grooves of a typical CW correlated with the MFA, which was ca. 35° in juniper and spruce CW. Also, the crystal length of cellulose in the cell walls of juniper is very similar to that of CW spruce. By contrast, it resembles to NW of almost any other wood species concerning the carbohydrate composition and especially the distribution of lignin and cellulose (Agarwal 2006; Gierlinger and Schwanninger 2006; Schmidt et al. 2009; Gierlinger et al. 2010; Hänninen et al. 2011b). The small amount of galactose, that is usually abundant in CW as (1–4)- β -galactan, indicated that juniper is closer to NW in its carbohydrate composition. As the wood densities of juniper and spruce CW are higher than the density of NW spruce, it can be concluded that the low MOE of juniper is due to high MFA.

The similarities and dissimilarities of juniper wood with CW can partly explain several of its peculiar material properties. Elucidation of the enzymatic mechanisms or genetics of the partly CW-like character of juniper could help understand the formation of reaction wood, in general, and the peculiarities of this interesting wood.

Acknowledgments

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